Development of the in vitro micronucleus test for exposure to whole aerosol from cigarettes and heated tobacco products

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Introduction

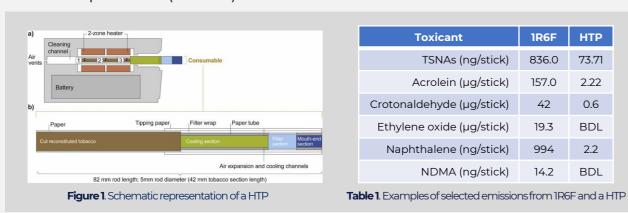
In the genotoxic assessment of tobacco products, traditional approaches involve capturing the two phases of the aerosol, such as pad-collected total particulate matter (TPM) and/or gas-vapour phase (GVP) in solvents and applied to cell cultures. This approach may under-represent the interactions of the whole aerosol (WA). The exposure of cells to WA at the air-liquid interface (ALI) mitigates the need for this fractionation.

The aim of this study was to investigate the potential of WA from a reference cigarette (1R6F) and a Heated Tobacco Product (HTP) to induce micronuclei in cells exposed at the ALI.

Methodology

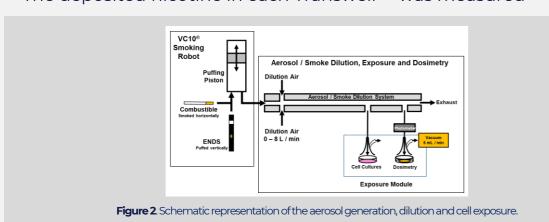
Test Articles

The 1R6F reference cigarette and a HTP were used in this study. A schematic of the HTP consumable and heating device are shown in Figure 1. Analytical chemistry demonstrates HTPs emit lower levels of toxicants per stick (Table 1)



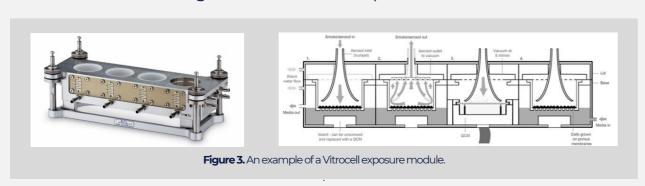
Aerosol Generation

- The Health Canada Intense (HCI; 55 mL puff volume, taken over 2 seconds, every 30 seconds, 100% vent blocking) regime was used for 1R6F
- The HTP was puffed using the same regime, but the ventilation holes
- The aerosols generated on the Vitrocell VC10 were diluted with fresh air (diluting airflow) to achieve the required aerosol dilution (Figure 2)
- Diluting airflows of 2 and 6 L/min were trialled with 1R6F, but only 6 L/min was used for the HTP
- The aerosol was then passed over the cells in Transwells™ at a vacuum rate of 5 mL/min (Figure 3)
- The deposited nicotine in each Transwell™ was measured



In vitro micronucleus assay

- V79 cells were seeded on 24 mm Transwells™ at 6 x10⁴ cells/mL
- Transwells™ incubated for 24 h to achieve approximately 50% confluency
- Prior to aerosol exposures, the apical media was removed from the surface of the cells, and the Transwells™ placed in the Vitrocell exposure modules and exposed for the required number of puffs
- Following exposure, Transwells™ were incubated for 24 h and analysed for micronuclei induction using Litron MicroFlow kits and cytotoxicity using RPD
- Air was used as the negative control and the positive control was MMC

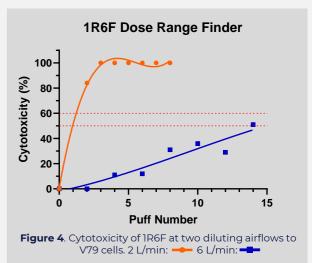




Results

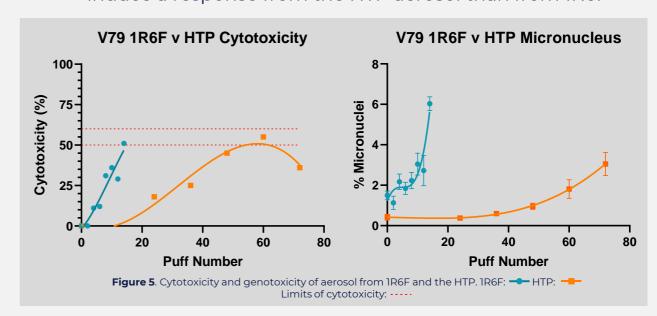
Diluting airflow optimisation

- With 1R6F WA, the 2 L/min diluting airflow was too cytotoxic, and no analysable concentrations were obtained
- The 6 L/min diluting airflow provided the optimal cytotoxic response (Figure. 4)

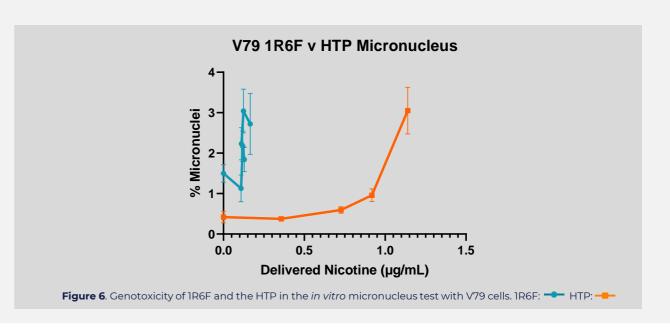


Micronuclei Induction

- The positive control, MMC, induced dose related increases in micronuclei that were considered positive responses (data not shown)
- 1R6F and the HTP aerosols induced dose related decreases in cell viability to 50-60%, and also induced dose related increases in micronuclei relative to the vehicle (air) control (Figure 5), but:
 - The HTP aerosol was 3 times more concentrated than 1R6F
 - No micronuclei were induced when HTP aerosol was tested at the same number of puffs as 1R6F
 - A significantly higher number of puffs was needed to induce a response from the HTP aerosol than from 1R6F



• When the dose was expressed as deposited nicotine, reduced genotoxicity of the HTP was still observed (Figure 6)



Conclusion

It is feasible to adapt submerged cell cultures to air-liquid interface exposures, a more biologically relevant approach than using separate assays for particulate and gas phase fractions, for gases and whole aerosols

The HTP was less genotoxic than 1R6F cigarette, on a puff-by-puff basis, and when the delivered dose was expressed as units of nicotine

This further supports the role of HTPs in the tobacco harm reduction agenda

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