

# Characterisation of a Vitrocell® VC 10 exposure system using dose tools and biological responses *in vitro*

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## Introduction

Traditional smoke exposure techniques are based on the particulate phase of tobacco smoke and not the complete whole smoke aerosol. The particulate phase of tobacco smoke makes up approximately 5-10% whereas the vapour phase makes up the remaining 90-95% (v:v)<sup>1</sup>. To overcome these challenges, whole smoke exposure systems have been developed that expose cell cultures to diluted tobacco smoke and capture the full interactions of both smoke phases.

## Aim

To characterise a Vitrocell® VC 10 smoke exposure system using a battery of tests. We achieved this by measuring smoke deposition<sup>2</sup> in relationship to airflow (L/min); vapour phase dilution using a known smoke marker carbon monoxide (CO). Finally we assessed biological responses using the Ames and Neutral Red uptake (NRU) assays.

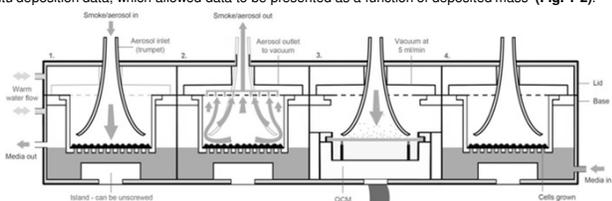
## Materials and Methods

### Cigarette Smoke Generation

Smoke was generated using a Vitrocell® VC 10 Smoking Robot, (Vitrocell® Systems, Waldkirch, Germany). Dilutions were achieved by diluting smoke in air (L/min), with a vacuum of 5ml/min/well. 3R4F cigarettes were smoked to the ISO smoking regime (35ml puff over 2 seconds, once a min - ISO 3308:1991).

### Measurement of Particulate Dose

Four quartz crystal microbalances (QCM) were installed into a 6PT-CF Vitrocell® exposure module to assess particulate dose<sup>3</sup>. During biological exposure, a QCM, located in position 4, recorded real-time *in situ* deposition data, which allowed data to be presented as a function of deposited mass (Fig. 1-2).



**Figure 1:** Schematic cross-section of the 6PT-CF mammalian exposure module depicting how smoke is delivered to cell culture inserts at the air-liquid interface (ALI) and the QCM module<sup>3</sup>. A similar setup, without media, was used for Ames agar plates.

### Measurement of Carbon Monoxide Concentration

CO concentrations were determined via the analysis of the diluted cigarette smoke, collected from exhaust in Douglas gas bags, using a Signal® 7000-FM gas analyser (Fig. 3).

### NRU Assay

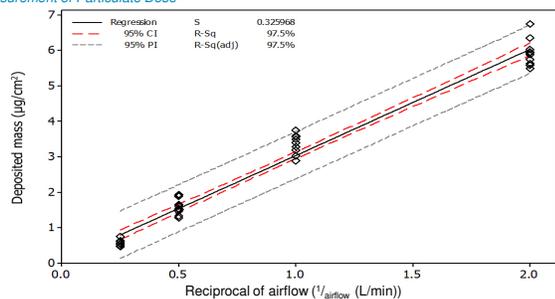
BalB/c 3T3 cells were seeded into 24 mm Transwells® and maintained in culture for ~24 h prior to exposure to achieve near-confluent monolayers. Cells were exposed at the ALI to freshly generated cigarette smoke. Following exposure (184 min), cells were assessed for cytotoxicity in the NRU assay (Fig. 4).

### Ames Assay

*Salmonella typhimurium* (YG1042) were plated onto 35 mm Vogel-Bonner agar plates in the presence of 10% S9 using a spread plate methodology. Bacterial agar plates were smoke exposed for 24 min, incubated at 37°C for 3 days and revertant colony numbers determined by automated scoring (Fig. 5).

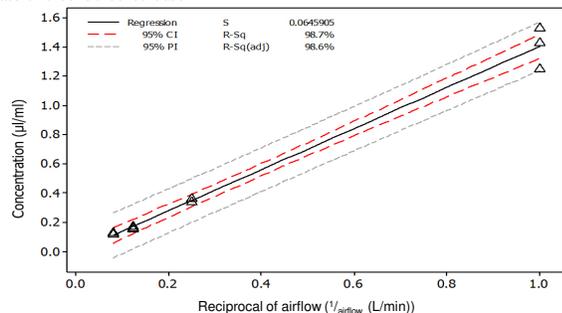
## Results

### Measurement of Particulate Dose



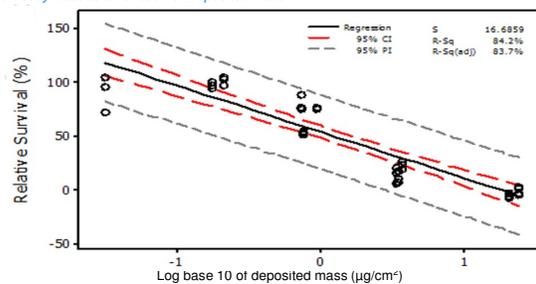
**Figure 2:** Measurement of deposited particulate mass from a 24 minute smoke exposure at four smoke dilutions (Airflows 0.5, 1.0, 2.0 and 4.0 L/min). Results are presented as a reciprocal of the airflow (1/airflow (L/min)) and are based on three independent experiments. A regression fit correlation of  $R^2 = 0.975$  was observed with 95% confidence intervals (red dash) and probability intervals (grey dash).

### Measurement of CO Concentration



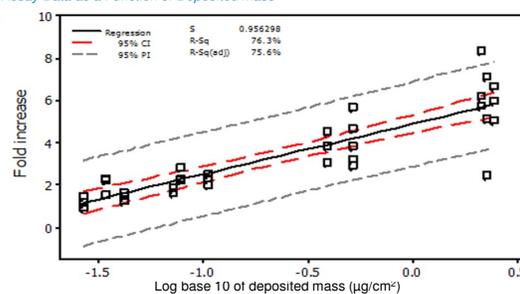
**Figure 3:** Measurement of CO concentrations following a 16 minute smoke exposure at four smoke dilutions (Airflows 1.0, 4.0, 8.0 and 12.0 L/min). Results are presented as a reciprocal of the airflow (1/airflow (L/min)) and are based on three independent experiments. A regression fit correlation of  $R^2 = 0.975$  was observed with 95% confidence intervals (red dash) and probability intervals (grey dash).

### NRU Assay Data as a Function of Deposited Mass



**Figure 4:** NRU assay relative survival data (v. air control) presented as a function of deposited mass (obtained *in situ* of exposure) on a log scale with confidence intervals (red dash) and probability intervals (grey dash) of 95%. A correlation of  $R^2=0.84$  was observed between deposited mass and cytotoxicity. Results are based on three independent experiments. Airflows = 1.0, 4.0, 8.0 and 12.0 L/min.

### Ames Assay Data as a Function of Deposited Mass



**Figure 5:** Ames assay fold increase data presented as a function of deposited mass (obtained *in situ* of exposure) on a log scale with confidence intervals (red dash) and probability intervals (grey dash) of 95%. A correlation of  $R^2=0.76$  was observed between deposited mass and mean fold increase in revertant numbers. Results are based on three independent experiments. Airflows = 1.0, 4.0, 8.0 and 12.0 L/min.

Airflow (L/min)	Reciprocal of airflow (1/airflow (L/min))	NRU Correlation of $R^2 = 0.847$		AMES Correlation of $R^2 = 0.763$		
		Mean deposited mass for NRU exposure ( $\mu\text{g}/\text{cm}^2 \pm \text{STDV}$ )	Mean % relative cell survival $\pm \text{STDV}$	Mean deposited mass for Ames exposure ( $\mu\text{g}/\text{cm}^2 \pm \text{STDV}$ )	Mean revertant fold increase $\pm \text{STDV}$	Mean total revertants $\pm \text{STDV}$
1.0	1.000	22.8 $\pm$ 1.7	-2.5 $\pm$ 3.3	2.30 $\pm$ 0.14	5.9 $\pm$ 1.6	78.6 $\pm$ 20.6
4.0	0.250	3.5 $\pm$ 0.1	16.7 $\pm$ 7.4	0.50 $\pm$ 0.10	4.0 $\pm$ 0.9	53.1 $\pm$ 9.6
8.0	0.125	0.8 $\pm$ 0.1	69.9 $\pm$ 13.0	0.09 $\pm$ 0.02	2.2 $\pm$ 0.4	30.2 $\pm$ 4.1
12.0	0.080	0.1 $\pm$ 0.1	96.8 $\pm$ 10.1	0.03 $\pm$ 0.01	1.6 $\pm$ 0.5	21.2 $\pm$ 5.0

**Table 1:** Raw data values including airflows, reciprocal of airflows, correlations, standard deviation between experiments, relative survival, mean revertant and fold increase numbers and *in situ* obtained deposited mass.

## Conclusions

- Correlations between airflow (L/min) and deposited mass and CO were observed, adding confidence to the exposure system:
  - Deposited mass correlation with airflow,  $R^2 = 0.97$
  - CO concentration correlation with airflow,  $R^2 = 0.98$
- The study demonstrated consistent responses using 2 independent biological assays for the assessment of cigarette smoke cytotoxicity (NRU) and mutagenicity (Ames)
  - Biological results have been presented as a function of deposited mass obtained *in situ* of exposure, giving our data a gravimetric measure. Correlations with deposited mass for both systems were observed:
    - NRU correlation with deposited mass,  $R^2 = 0.84$
    - Ames correlation with deposited mass,  $R^2 = 0.76$
- As CO is only one smoke marker and may not be representative of the complete aerosol, additional thoughts should be given to alternative gas phase dose tools
- It is likely that both fractions of cigarette smoke play an important role in biological responses and therefore it is important to characterise these within an *in vitro* system
- This study has increased our knowledge and working understanding of the VC 10 exposure system, but further work is required to fully characterise this system

## References

- Clunes, L., Bridges, B., Aloisi, N., Tarran, R. *In vivo* versus *in vitro* airway surface liquid nicotine levels following cigarette smoke exposure. *Journal of Analytical Toxicology* 2008; 32: 201-207.
- Adamson, J., Hughes, S., Azzopardi, D., McAughy, J., Gaca, M. Real-time assessment of cigarette smoke particle deposition *in vitro*. *Chemistry Central Journal* 2012; 6:98.
- Adamson, J., Thorne, D., Dalrymple, A., Dillon, D., Meredith, C. Cigarette smoke deposition in a Vitrocell® exposure module: real-time quantification *in vitro* using quartz crystal microbalances. *Chemistry Central Journal* 2013; 7:50