**Application of Next Generation In Vitro Approaches for the Assessment of E-Cigarettes**

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**Introduction**

Electronic cigarette (e-cigarette) use has increased at a rapid pace over the last decade. As these products continue to evolve to meet consumer demand, there is an increased requirement for the toxicological evaluation of e-cigarettes and e-liquids.

Using a series of classical and next generation in vitro test methods, we have assessed the toxicological and biological effects of a prototype e-cigarette compared to a commercially reference cigarette (3R4F or 1R6F).

**Materials and Methods**

Schematic drawing of novel prototype e-cigarette. a) e-liquid; b) distiller technology; c) electronics and battery; d) device body.

**Test matrix preparation**

Health Canada Intense for 3R4F/1R6F reference cigarette and CORESTA Recommended Method No81 for prototype e-cigarette puffing regimens were used to generate three different test matrices for in vitro assessment:

- Total Particulate Matter (TPM) – trapped on a Cambridge filter pad and eluted at 24 mg/mL (3R4F) and 150 mg/mL (e-cigarette) in DMSO. TPM was diluted in cell culture medium to treat cells in submerged culture.
- Whole aerosol (WA) – cells directly exposed to aerosol at the air-liquid interface.
- Aerosol aqueous extract (AqE) – aerosol bubbled through 20 mL cell culture medium in a glass impinger. 8 puffs were used for 3R4F, while 24 puffs of e-cig were used. AqE was diluted in cell culture medium to treat cells in submerged culture.

**Classical regulatory toxicology**

* Ames bacterial reverse mutation assay*

TPMs were tested using five S. typhimurium strains: TA98, TA100, TA1535, TA1537 and TA102 + metabolic activation (S9) following OECD 471. For WA exposures, S. typhimurium tester strains TA98, TA100, TA1535, TA97 and TA102 using a modified air-agar methodology.

* In vitro micronucleus (IVMN)*

TPMs were assessed following OECD 487, with short 3hrs exposures (± S9) and longer 24hrs -S9 exposures.

* Mouse lymphoma assay (MLA)*

TPMs were assessed following OECD 490, with short 3hrs + S9 exposures and longer 24hrs - S9 exposures.

* Neutral red uptake (NRU) cytotoxicity assay*

Cytotoxicity of TPMs were assessed using BALB/c 3T3 mouse fibroblasts. WA cytotoxicity was assessed using human bronchial epithelial cells (H292) exposed at the air-liquid interface (ALI).

* Bhas cell transformation assay*

The potential of TPMs to induce tumour development was evaluated using the Bhas 42 cell transformation assay promoter protocol.

**Next generation toxicology**

**Multiparametric analysis using high-content screening (HCS) approaches**

The Cellomics ArrayScan VT platform was used to assess 10 endpoints including cell health markers, DNA damage and oxidative stress, in normal human bronchial epithelial cells (NHBEs) after 4hrs or 24hrs exposures to TPM.

**Luciferase-based reporter gene assay to assess oxidative stress**

Antioxidant response element (ARE) transpositional activation in stably transfected H292 cells were assessed after 6hrs and 24hrs exposures to TPM.

**Endothelial cell migration assay**

Human umbilical vein endothelial cells (HUVEC) were grown to confluency, “wounded” using a pipette tip and exposed to various AqE for 24hrs in 24 well ImageLock plates. Cell migration was quantified by measuring the closure of the “wound” using an Incucyte time lapse video camera and software at hourly intervals.

**Systems toxicology: transcriptomics and proteomics**

MucilAir human 3D primary airway epithelial cultures were exposed to 1R6F or e-cigarette aerosols at the ALI. After exposure, cells were incubated for 24hrs or 48 hrs. The cell lysates used for RNA-seq transcriptomic analysis and the air-surface liquid (ASL) was collected for LC-MSMS proteomics analysis.

**Results**

### Classical regulatory toxicology

**Cytotoxicity TPM**

**AMES TA98 TPM**

**AMES TA100 TPM**

**IVMN (3hrs +S9)**

**Bhas cell transformation**

**MLA(3hrs +S9)**

**Ames TA98 WA**

**Ames TA100 WA**

A battery of genetic toxicology assays including cytotoxicity, mutagenicity, clastogenicity and cell transformation were employed to test both TPM and WA matrices generated from cigarette smoke and a prototype e-cigarette.

** Compared to cigarette smoke, the prototype e-cigarette showed significantly reduced activity in all assays, even at extreme doses.**

### Next generation toxicology

#### Whole aerosol cytotoxicity

WA cytotoxicity assessment demonstrated that 3R4F produced a decrease in cell viability, resulting in complete cytotoxicity after just 7 puffs of undiluted aerosol. The novel e-cigarette induced significantly less cytotoxicity at up to 1,000 puffs and at comparable and higher levels of nicotine delivered to the cells.

#### Reporter gene assay

**Activation of the H230-ARE-Luc2P following exposure to TPM from 3R4F and a novel e-cigarette. Data showed a mean fold change in response normalized to the vehicle control (0.83% DMSO). 3R4F caused ARE activation at both time points. In contrast, the novel prototype e-cigarette was negative in the ARE assay.**

### RNA-seq differential gene expression

Transcriptomics profile of MucilAir™ after exposure to 1R6F smoke or e-cigarette aerosol. 1R6F exposure triggered the differential expression of 5603 genes at 24 hrs, 2180 genes at 48 hrs, and 6045 genes for the time-adjusted contrast. No differently expressed genes were detected when comparing e-cigarette exposure RNA-seq data to the air control. Volcano plots were generated using a fold change threshold of [FC] > 1.5, and an adjusted p-value significance threshold of p<0.05.

**Targetted and untargetted proteomics**

Targeted and global proteomics profile of ASL collected from MucilAir exposed to smoke or e-cigarette aerosol. (A) Targetted MRM MLL-based quantification of selected ASL proteins relative to air control after 1R6F or prototype e-cigarette exposure. At 24 hrs or 48 hrs post-exposure. (B) Label-free quantification of total ASL proteins of MucilAir exposed to 1R6F (top panel) or e-cigarette) at 24 and 48 hrs post-exposure at [FC] > 1.5 and p<0.05. 1R6F smoke exposure treatments with a recovery time of 24hrs and 48 hrs were found associated with a total of 16 and 17 differentially abundant proteins, respectively. At the same significance threshold, e-cigarette aerosol treatments with a recovery time of 24hrs and 48 hrs were found associated with only 1 and 2 differentially abundant proteins, respectively.

**Conclusions**

Compared to cigarette smoke, the prototype e-cigarette showed significantly reduced activity in all assays, even at extreme doses.

Next generation toxicological approaches can be used as part of an in vitro testing strategy for e-cigarettes, in concert with classical toxicological testing.

The results show this novel e-cigarette has the potential to be a reduced risk product though further longer-term studies would be required to substantiate this potential.